Turkish Journal of Chemistry

Volume 41 | Number 2

Article 1

1-1-2017

Isocoumarins and 3,4-dihydroisocoumarins, amazing natural products: a review

AISHA SADDIQA

OSMAN ÇAKMAK

MUHAMMAD USMAN

Follow this and additional works at: https://journals.tubitak.gov.tr/chem

Part of the Chemistry Commons

Recommended Citation

SADDIQA, AISHA; ÇAKMAK, OSMAN; and USMAN, MUHAMMAD (2017) "Isocoumarins and 3,4-dihydroisocoumarins, amazing natural products: a review," *Turkish Journal of Chemistry*: Vol. 41: No. 2, Article 1. https://doi.org/10.3906/kim-1604-66 Available at: https://journals.tubitak.gov.tr/chem/vol41/iss2/1

This Article is brought to you for free and open access by TÜBİTAK Academic Journals. It has been accepted for inclusion in Turkish Journal of Chemistry by an authorized editor of TÜBİTAK Academic Journals. For more information, please contact academic.publications@tubitak.gov.tr.



Turkish Journal of Chemistry

http://journals.tubitak.gov.tr/chem/

Review Article

Turk J Chem (2017) 41: 153 – 178 © TÜBİTAK doi:10.3906/kim-1604-66

Isocoumarins and 3,4-dihydroisocoumarins, amazing natural products: a review

Aisha SADDIQA^{1,*}, Muhammad USMAN², Osman ÇAKMAK³

¹Department of Chemistry, Faculty of Natural Sciences, Government College Women University, Sialkot, Pakistan ²Department of Chemistry, Government College of Science, Lahore, Pakistan ³Department of Nutrition and Dietetics, School of Health Sciences, İstanbul Gelişim University, Avcılar, İstanbul, Turkey

Received: 23.04.2016 •	Accepted/Published Online: 10.09.2016	٠	Final Version: 19.04.2017
-------------------------------	---------------------------------------	---	----------------------------------

Abstract: The isocoumarins are naturally occurring lactones that constitute an important class of natural products exhibiting an array of biological activities. A wide variety of these lactones have been isolated from natural sources and, due to their remarkable bioactivities and structural diversity, great attention has been focused on their synthesis. This review article focuses on their structural diversity, biological applications, and commonly used synthetic modes.

Key words: Isocoumarin, synthesis, natural product, biological importance

1. Introduction

The coumarins 1 are naturally occurring compounds having a fused phenolactone skeleton. Coumarin 1 was first extracted from *Coumarouna odorata* (tonka tree).¹ The isocoumarins 2 and 3,4-dihydroisocoumarins 3 are the isomers of coumarin 1. A number of substituted isocoumarins have been found to occur in nature; however, the unsubstituted isocoumarins have not been observed to occur naturally. Furthermore, sulfur, selenium, and tellurium analogues 4a-4c have also been known since early times (Figure 1).

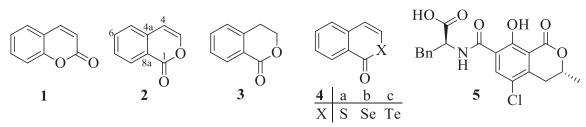


Figure 1. Some naturally occurring isocoumarins.

The isocoumarins and their analogues occur in nature as secondary metabolites (i.e. produced by living beings in response to external stimuli) of plants and lower microorganisms. A few isocoumarins are also extracted from insect pheromones and venom. These lactones are structural subunits of several natural products and serve as useful intermediates in the synthesis of different heterocyclic molecules. The isocoumarins have been found to exhibit beneficial (e.g., antitumor, antileukemic, antiviral, and antimicrobial²) as well as toxic biological activities; for example, ochratoxin A **5** is a potent mycotoxin produced by *Aspergillus* and *Penicillium* species, which is hepatotoxic, nephrotoxic, teratogenic, and carcinogenic in animals.³

^{*}Correspondence: aashe06@gmail.com

Due to the pharmacological and biochemical properties and the therapeutic applications of isocoumarins and 3,4-dihydroisocoumarins, research concerning the isolation and syntheses of isocoumarins has caught the attention of many organic chemists, which is reflected by the large number of review articles that have been published on isocoumarins. For example, Barry,⁴ Turner and Aldridge,⁵ Napolitano,⁶ Bin,⁷ and Saeed⁸ published reviews about isocoumarins and 3,4-dihydroisocoumarins.

1.1. Nomenclature

The name "isocoumarin" is derived from the fact that these compounds are isomers of coumarins **3**. The IUPAC names of isocoumarins **1** and their 3,4-dihydroanalogues **2** are 1H-2-benzoxin-1-ones and 3,4-dihydro-1H-2-benzoxin-1-ones, respectively. In the literature no proper nomenclature exists for isocoumarins and 3,4-dihydroisocoumarins. Generally, the trivial names derived from specific or generic names of fungal or plant sources are used for naturally occurring isocoumarins and their 3,4-dihydroanalogues.⁸ Names such as alternariol (*Alternaria* sp.), peniolactol (*Peniophora sanguinea*), cladosporin (*Cladosporium* sp.), and homalicine (*Homalium zeylancum*) are common examples of the names derived from genera and mellein (*Aspergillus melleus*), ustic acid (*A. ustus*), and duclauxin (*P. duclauxi*) are examples of names derived from specific names.

Most of the trivial names of isocoumarins end in suffixes such as -in, -ol, -one, -ide, -oic acid, or anhydride depending on the nature of the functional group present. Some examples are artemidin, altenuisol, oospolactone, agrimonolide, ustic acid, β -callatolic acid, lamellicolic anhydride, and naphthalic anhydride.

2. Pharmacological importance

The isocoumarins and 3,4-dihydroisocoumarins are an important class of naturally occurring lactones isolated from different bacterial strains, molds, lichens, and plants. They show a wide range of biological activities, ranging from antibacterial to antitumour.^{9–11} Significant work has been published about their biology and chemistry. Some of the selected biological activities are discussed here.

Amicoumacin A **6** and C **7** have been found to show antiulcer, antibacterial, and antiinflammatory activities.¹² Baciphelacins **8** have good potential for the treatment of bacterial and viral infections.^{13,14} Among dihydroisocoumarins, PM-94128 **9**¹⁵ and Y-05460M-A **10**¹⁶ have been found to exhibit antiulcer activity in addition to antibacterial and antitumor activities (Figure 2).

The activity of PM-94128 **9** was examined against four different tumor cell lines including P-388 (lymphoid leukemia), A-549 (human lung carcinoma), HT-29 (human colon carcinoma), and MEL-28 (human melanoma) in the 50 nM activity range.¹⁷ Amicoumacin Sg17-1-4 **11** isolated from a marine fungus, *Alternaria tenuis* Sg17-1, shows cytotoxic activity against HeLa cell lines (Figure 2).¹⁸

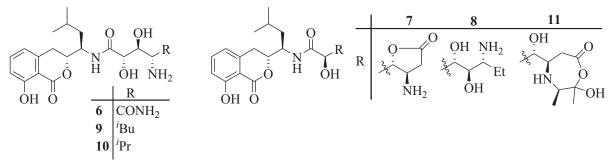


Figure 2. 3,4-Dihydroisocoumarins with antibacterial and antitumor activities.

A series of structurally related isocoumarins known as A1-77s are a small family of antibiotics isolated from a culture broth of *Bacillus pumilus* A1-77 (found in the gut of *Coenagrion* dragonfly larvae and also produced by *Nocardia jinanesis*) (Figure 3).¹⁹ The structural feature of A1-77s comprises a dihydroisocoumarin moiety connected to different acyl hydroxy amino acid chains. It is the variation in the amino acid chain that results in various members of the family.^{20,21} These compounds possess a broad range of pharmacological properties including antibacterial, antiinflammatory, antiulcer, gastroprotective, and anti-*Helicobacter pylori* activities.^{12,22-24} However, they are famous for their remarkable gastroprotective activity.¹⁹ The family members of the A1-77s include compounds AI-77-A **12**, -B **13**, -C **14**, -D **15**, -F **16**, and -G **17**, which vary in their acyl hydroxy amino acid chains (Figure 3).¹²

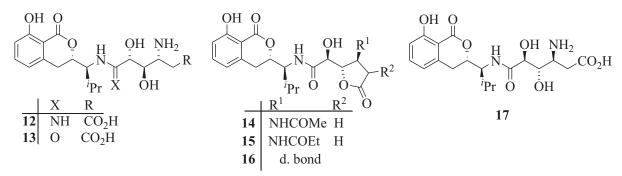


Figure 3. A series of structurally related A1-77 isocoumarins.

AI-77-B is the most abundant compound of the amicoumacin family. It is also known as amicoumacin B 13, which is a major product of the fermentation process and has been found to exhibit potent gastroprotective β -amino acid 18.²⁵ Amicoumacin B 13 shows antiinflammatory (rats), antiulcer (human stomachs), and herbicidal (*Lemna*) activities and is also used as an acaricide.²⁶ Besides its unique structure and its characteristic biological activity, its therapeutic potential is limited because of poor oral absorption properties. As a result, structural modifications and synthetic studies of AI-77-B have attracted a great deal of attention from synthetic chemists.^{27–30}

Bergenin 19,³¹ isolated from *Flueggea microcarpa* and *Flueggea virosa*, and an isocoumarin coriandrin 20,³² isolated from *Coriandrum sativum*, have been found to show anti-HIV activity (Figure 4). In addition to anti-HIV, bergenin 19 also possesses antiulcer³³ and antihepatotoxic activity.³⁴ It was observed that 19, extracted from the aboveground parts of *Flueggea virosa*, was proved to have good potential to treat cardiac arrhythmias.³⁵ The one isolated from *Flueggea microcarpa* showed antifungal activity against several plant-pathogenic fungi.³⁶

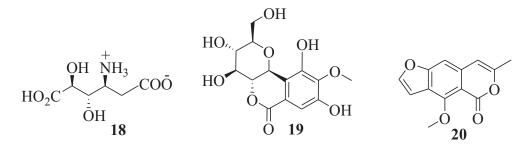


Figure 4. Structures of β -amino acid 18, bergenin 19, and coriandrin 20.

The 9,10-dihydroxy-5,7-dimethoxy-1H-naphtho-(2,3c)pyran-1-one **21** (common name: paepalantine) is extracted from *P. bromelioides* and shows intense cytotoxic activity in the McCoy cell line (Figure 5). The paepalantine molecule is more lipophilic than other isocoumarins because hydroxyls at positions C⁹ and C¹⁰ form an intramolecular bridge of a hydrogen bond. It has been found that the rate of cytotoxicity depends on the presence and position of hydroxyl group in the isocoumarin framework.³⁷ Thunberginol B **22a–22b**, naturally occurring isocoumarins, also show anticancer effect (Figure 5). In addition, thunberginol B has been found to have antiallergic, antimicrobial, antioxidant, and choragic activities.³⁸

Isocoumarin derivatives 6,8-dihydroxy-4-acetyl-isocoumarin **23** and 6,8-dihydroxy-4-(1-hydroxyethyl)isocoumarin **24** are effective angiogenesis inhibitors (Figure 5).³⁹ A group of 3-carboxyisocoumarins **25a**–**25c** showed antiallergic effects (Figure 6).⁴⁰

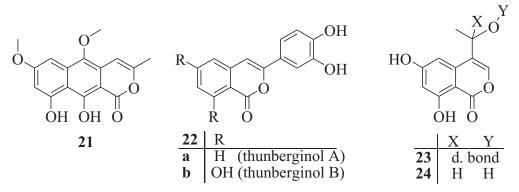


Figure 5. Structures of paepalantine 21, thunberginol B 22, and 6,8-substituted 3,4-dihydroisocoumarin 23-24.

Inhibitors are the chemical substance that reduces the activity of enzymes by blocking the active sites of enzymes. Serine proteases are essential inhibitors that play significant roles in various physiological processes such as blood coagulation, digestion, viral infection, fibrinolysis, and fertilization. They can also be lethal if they are uncontrolled. They can cause various diseases such as tumors, cerebral infection, emphysema, vascular clotting, arthritis, and bronchial inflammation. It is thus necessary to introduce a variety of selective inhibitors for the treatment of diseases related to serine proteases.⁴¹

The chloro- and amino-substituted isocoumarins, e.g., 3-bromoalkoxy-4-chloro-7-benzamidoisocoumarins **26**, are well-known compounds for the development of uncharged inhibitors of urokinase-type plasminogen activator (uPA) (Figure 6). They have important contributions to the extracellular proteolytic events associated with tumor cell growth, migration, and angiogenesis.⁴² The aminoalkoxy- and guanidino-substituted isocoumarins **27** have also been found as powerful inhibitors of blood coagulation serine proteases (Figure 6).⁴³

Various derivatives of isocoumarins and 3,4-dihydroisocoumarins were screened for their antibiotic activity. Among the substituted isocoumarins, the 8-hydroxylisocoumarins **28**, 6,8-dihydroxy-3-(4-hydroxyphenyl)isocoumarins **29**, and 8-dihydroxy-3-(3,4-dihydroxyphenyl)-isocoumarins **30** possess strong antibiotic effects (Figure 7). The isocoumarins **31–33** have also been found effective against various strains of gram-positive and gram-negative bacteria such as *Escherichia coli*, *Pseudomonas aeruginosa*, *Shigella boydii*, *Salmonella* serovar Typhi, and *Bacillus cereus* (Figure 7).⁴⁴

The 3,4-dihydroisocoumarins 34-35 were examined for antibacterial effects against different grampositive and gram-negative bacterial strains (Figure 8).⁴⁵

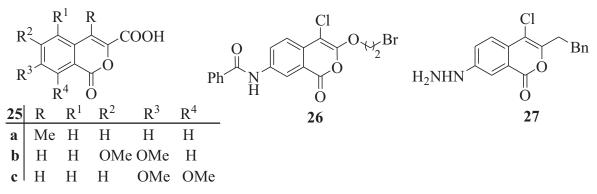


Figure 6. Structures of antiallergic and inhibitor isocoumarins 25a-25c and 26-27.

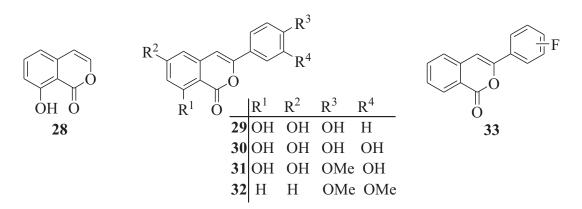


Figure 7. Hydroxy isocoumarins possessing strong antibiotic effects.

Some other derivatives of 3,4-dihydroisocoumarins, such as 3-(3',4'-dimethoxyphenyl)-3,4-dihydroisocoumarin and 3-(3',4'-dihydroxyphenyl)-3,4-dihydroisocoumarin **36a**–**36b**, showed moderate effects when tested in vitro for antibacterial activity (Figure 8).⁴⁶

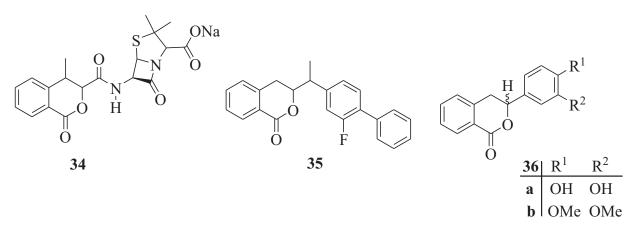


Figure 8. Antibacterial 3,4-dihydroisocoumarins.

Malaria is a potentially fatal blood disease of tropical climate areas. It is caused by eukaryotic protists of the genus *Plasmodium*. *Plasmodium* is present in the body of humans and an animal host, the *Anopheles*

mosquito, and is transferred to human blood by the bite of infected *Anopheles* mosquitoes. The most common drug prescribed for malaria patients is chloroquine, but some derivatives of 3,4-dihydroisocoumarin, mullein **37a–37d**, produced by *Botryosphaeria rhodina*, an endophytic fungus, show effective antimalarial activity (Figure 9).⁴⁷ Some isocoumarin derivatives, 8-hydroxy-6-methoxy-3-pentyl-1H-isochromen-1-one **38a–38b**^{48,49} and halorosellins **39**,⁵⁰ isolated from the bark and stem of *Tessmannia densiflora* and *Halorosellinia oceanica*, respectively, also showed antimalarial activity (Figure 9).

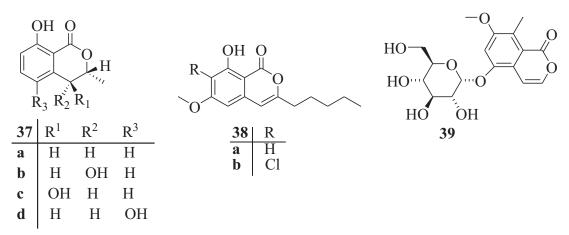


Figure 9. Structures of antimalarial molecules.

Ceratocystis fimbriata, a fungus, is a source of isocoumarins 40a-40d that are known for their phytotoxic activity on leaves of coffee trees, and also they are responsible for fruit withering with trunk canker in adult coffee trees (Figure 10).⁵¹

Other derivatives of 3,4-dihydroisocoumarin **41a–41b** extracted from the fungus *Ceratocystis ulmi* inhibit the growth of rice seedlings and lesions on the leaves of pear trees (Figure 10).⁵²

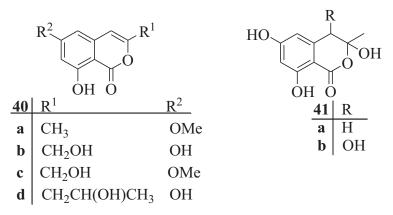


Figure 10. Structures of isocoumarins 40 and dihydrocoumarins 41.

Mutation is a variation in the fundamental coding series of the hereditary material, which in most plants and animals is DNA, but in a few viruses is RNA. It occurs by new genetic recombinations of nitrogenous bases present in the hereditary material of organisms. Mutations have proved to be fatal and were found to cause various hereditary diseases. There are some other processes that create change in the genotype of an organism but are not referred to as mutations, and these include combinations of chromosomes in the offspring, artificially induced recombinations, or the introduction of new genetic material into an organism.

Paepalantine-9- α -D-glactopyranoside **42**, a glucose derivative extracted from *Paepalanthus bromelioides*,⁵³ and isocoumarins **43a**–**43b** have been found to exhibit mutagenic effects (Figure 11).^{53,54}

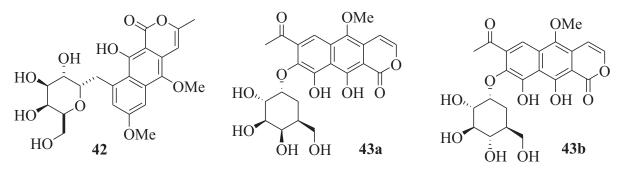


Figure 11. Glucose derivatives of isocoumarins.

The 3-(2-naphthyl)3,4-dihydroisocoumarin 44 was tested in vitro against fungal strains *C. albicans*, *F. solani*, *T. schoenleinii*, *A. niger*, *M. phaseolina*, and *P. boydii* and it was found active against all the fungal strains except *C. albicans* (Figure 12).⁵⁵ Monocerin 45 and its different analogues have been isolated from numerous fungal sources, such as *Drechslera monoceras* (Figure 12). They possess excellent antifungal properties.⁵⁶

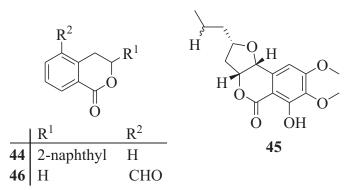


Figure 12. Structure of antifungal isocoumarines 44-45 and erythocentaurin 46.

Erythocentaurin **46** is isolated from *Enicostema hyssopifolium*, which is widely distributed in southern Pakistan. This plant is considered medicinally important is and used locally by the indigenous people as a remedy for malaria. In different regions of Pakistan, other species from the same family are used as digestive aids, as stomachic tonics, and for depurative, sedative, and antipyretic effects. Erythocentaurin **46** has also been found to be an active agent against serine proteases such as chymotrypsin and trypsin; these proteases are involved in the destruction of certain fibrous proteins.⁵⁷

The derivative of 3,4-dihydroisocoumarin **47** has antileukemic activity (Figure 13).⁵⁸ Isocoumarins **48a**–**48b** have been found useful for the cure of diseases associated with an abnormality in immunological regularity function or vascularization (Figure 13).⁵⁹ 3,4-Dihydroisocoumarin **49** has been found as trail pheromone in the hindgut of ants of various species of the genera *Formica* and *Lasius*.⁶⁰ A number of derivatives of isocoumarins are used as sweetners, ⁶¹ e.g., **50** (Figure 13).

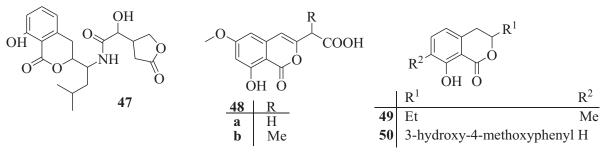


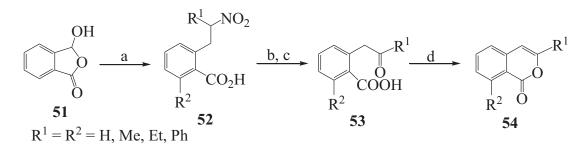
Figure 13. Structures of hydroxy coumarins 47-50.

The isocoumarins and 3,4-dihydroisocoumarins are naturally occurring lactones that display a wide range of biological and pharmacological activities $^{62-67}$ and serve as key intermediates in the synthesis of biologically active molecules. These are identified as highly attractive molecules in organic chemistry. $^{64-67}$ A wide spectrum of synthetic methods have been used for the synthesis of isocoumarins and 3,4-dihydroisocoumarins. 68 A number of new methods $^{69-79}$ are being developed and reported each year. Some of these methods provide the isocoumarins directly, whereas others lead to the 3,4-dihydroisocoumarins. Some of the most important high-yielding methods applicable to the synthesis of a large number of these compounds are reported below.

3. Synthetic approaches

3.1. Regiospecific synthesis of isocoumarins

Hauser et al.⁸⁰ reported the synthesis of isocoumarin **54** from phthalaldehydic acid **51** and nitroalkanes. The condensation of **51** afforded (nitroalkyl)benzoic acids **52** in good yield (70%–95%). The Nef reaction of **52** yielded ketoacid **53**, which upon intramolecular cyclization followed by dehydration yielded isocoumarin **54** (Scheme 1).



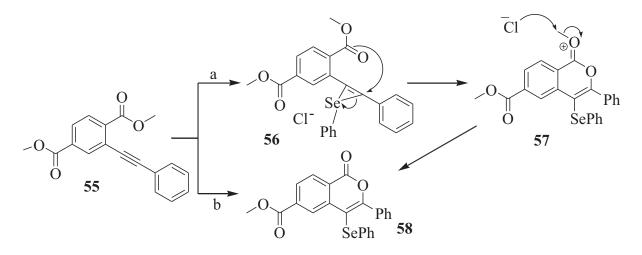
Reagent and conditions: a) RCH₂NO₂, Et₃N, DMSO; b) NaBH₄, DMSO; c) i. NaOH, ii. H₂SO₄, MeOH;

d) Ac₂O, EtOAc, H^+ .

Scheme 1. Synthetic scheme of isocoumarin 54.

3.2. Synthesis of isocoumarins via electrophilic cyclization

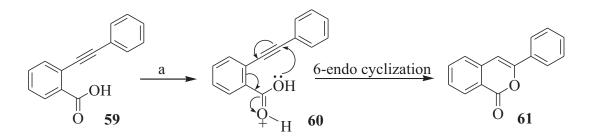
Yao and Larock⁷⁶ reported the synthesis of isocoumarins via electrophilic cyclization of o-(1-alkynyl)benzoates 55. A series of substituted isocoumarins 58 were synthesized in good yields under mild reaction conditions by the reaction of various o-(1-alkynyl)benzoates 55 with electrophiles such as ICl, I_2 , PhSeCl, and HI. The reaction proceeded through intermediates 56–57 (Scheme 2).



Reagent and conditions: **a**) PhSeCl; **b**) PhSeCl (1.2 eq), CH₂Cl₂, rt. **Scheme 2.** A representative of electrophilic cyclization for the synthesis of various isocoumarins.

3.3. Acid-catalyzed cyclizations of 2-(phenylethynyl)benzoic acid

Uchiyama et al. carried out the acid-catalyzed selective cyclization of enynecarboxylic acid **59** to isocoumarin **61** via intermediate **60** (Scheme 3).⁸¹



Reagent and conditions: a) TfOH or CF₃COOH.

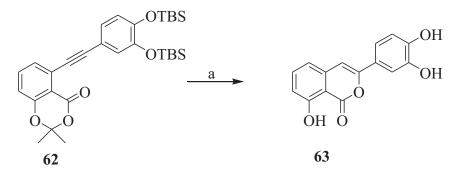
Scheme 3. Acid-catalyzed selective cyclization of an enynecarboxylic acid 59 to isocoumarin 61 via intermediate 60.

This strategy was applied for the synthesis of thunberginol A **63** from **62**, known for having miscellaneous biological applications, such as antimicrobial and antiallergic activities (Scheme 4).⁸¹⁻⁸⁵

3.4. Synthesis involving metals/metal ions/transition metal complexes

The literature shows that isocoumarins and 3,4-dihydroisocoumarins have been widely prepared in ways involving metalation at certain positions. Such strategies include lithiation and silylation.

Menashe et al. reported that diphenylacetylene **64** reacts with AcOH **65** in the presence of Ru-catalyst under reflux conditions to afford isocoumarin **66**. The mechanism of this transformation is still ambiguous; however, it was observed that the Ru-catalyst plays an important role in this reaction as the reaction does not proceed in the absence of this catalyst (Scheme 5).⁸⁶



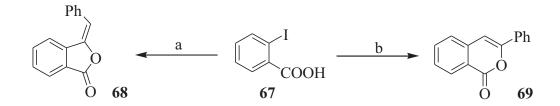
Reagent and conditions: a) TfOH, THF, reflux, 7 h.Scheme 4. Synthesis of thunberginol A 63 from 62.



Reagent and conditions: a) Ru₃(CO)₁₂

Scheme 5. Ru-catalyzed synthesis of isocoumarin 66 from diphenylacetylene 64.

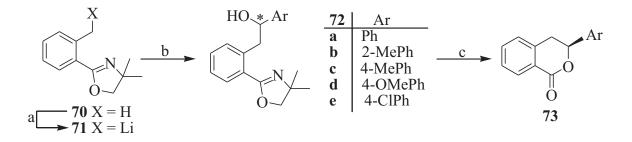
While o-iodobenzoic acid **67** was reacted with Cu-acetylides to yield 3-benzylidinephthalide **68** instead of the formation of isocoumarin, the same acid upon reaction with phenyl acetylene in the presence of a catalytic amount of Cu(I)-PPh₃ and K_2CO_3 under microwave atmosphere yielded isocoumarin **69** as a major product (Scheme 6).⁸⁷



Reagent and conditions: **a**) CuCCPh, C₅H₅N, reflux; **b**) HCCPh, Cu(I)-IPPh₃, K₂CO₃, microwave irradiation.

Scheme 6. Microwave-assisted synthesis of isocoumarin 69.

The oxazoline **70**, upon deprotonation with ^sBuLi followed by the addition of external chiral ligand (S)-2-(1-pyrrolidinylmethyl)pyrrolidine, yielded the lithiated species **71**. It was then treated with PhCHO to afford alcohol **72**, which upon further hydrolysis under mild conditions yielded isocoumarin **73** (Scheme 7).⁸⁸

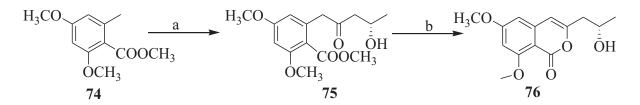


Reagent and conditions: a) Et₂O, -78 °C, ^sBuLi, (S)-2-(1-pyrrolidinylmethyl)pyrrolidine Li; b) PhCHO;

c) MeOTf, Et₂O, EtOH, reflux.

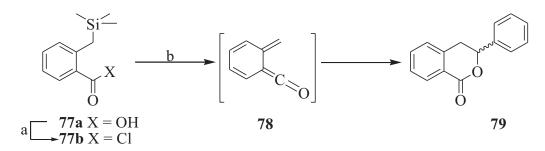
Scheme 7. Scheme for the synthesis of isocoumarin 73.

The methyl 2,4-dimethoxy-6-methylbenzoate **74** was lithiated with lithium diisopropylamide (LDA) and reacted with ethyl (S)-3-hydroxybutyrate to produce (S)-methyl-2-(4-hydroxy-2-oxopentyl)-4,6-dimethoxy benzoate **75**. The condensation of **75** with TsOH afforded 3,4-dihydroisocoumarin **76** (Scheme 8).⁸⁹



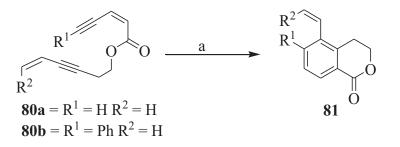
Reagent and conditions: a) LDA, ethyl (S)-3-hydroxybutyrate, HCl; b) TsOH, PhH, reflux.Scheme 8. Synthetic strategy for 3,4-dihydroisocoumarin 76.

The 2-(trimethylsilylmethyl)benzoyl chloride 77b upon desilylation followed by the reaction of ketene 78 with benzaldehyde yielded dihydroisocoumarins 79 (Scheme 9).⁹⁰



Reagent and conditions: a) SOCl₂; b) CsF. ArCHO.Scheme 9. A synthetic method for 3,4-dihydroisocoumarin 79.

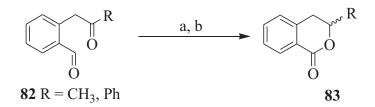
The single-step synthesis of 5,6-substituted 3,4-dihydroisocoumarins **81** was carried out by Kawasaki and coworkers via Pd-catalyzed intramolecular benzannulation reaction of bis-enynes **80** (Scheme 10).⁹¹



Reagent and conditions: a) Pd(PPh₃)₄, DPPF, Ph-Me, 80 °C

Scheme 10. The single-step synthesis of 5,6-substituted 3,4-dihydroisocoumarins 81 developed by Kawasaki and coworkers.⁹¹

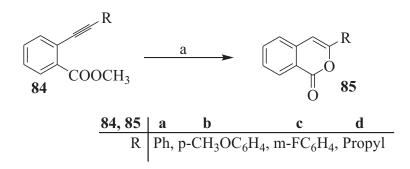
Suzuki et al. reported the oxidative lactonization of δ -ketoaldehydes **82** by exploiting an Ir-ligand bifunctional catalyst to afford coumarin derivatives. The intramolecular Tishchenko reaction of δ -ketoaldehydes afforded 3,4-dihydroisocoumarin **83** in good yields (Scheme 11).⁹²



Reagent and conditions: a) Ir-catalyst, rt; b) Tishchenko reaction.

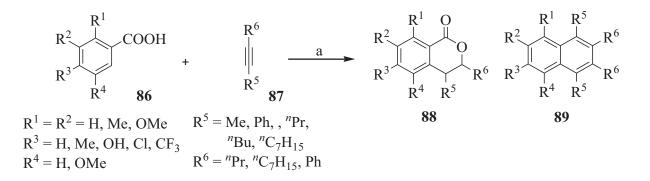
Scheme 11. Intramolecular Tishchenko reaction of δ -ketoaldehydes for the synthesis of 3,4-dihydroisocoumarin 83.

Marchal et al.⁹³ carried out Au(I)-catalyzed intramolecular cyclization of esters **84a–84d** to various alkylidene lactones **85**. The electronic effects of the R group and bulky substituents on the alkyne strongly modify the reactivity. The formation of isocoumarins from the cycloisomerization of o-alkynylbenzoic methyl esters is catalyzed by 10 mol% AuCl in the presence of 2 equivalents of H₂O. Under these conditions, several lactone rings **85a–85d** are formed in 60%–83% yield (Scheme 12).



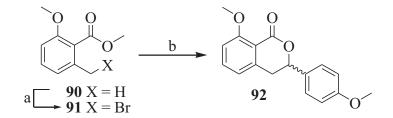
Reagent and conditions: a) AuCl (10 mol%), 2 eq. H₂O, MeCN, 50 °C, 24–48 h. Scheme 12. Synthesis of isocoumarins 85 by the cycloisomerization of o-alkynylbenzoic methyl esters.

Miura and coworkers⁹⁴ described the Rh-catalyzed direct oxidative coupling of benzoic acids **86** with internal alkynes **87** that leads to the formation of 6-membered lactones **88** as the major products and naphthalene derivatives **89** as by-products. The reaction of **86** with dialkylacetylenes proceeded efficiently to produce 3,4dialkylisocoumarins in good yields. Using unsymmetrical alkylphenylacetylenes, 4-alkyl-3-phenylisocoumarins **88** were predominantly formed in 84%–89% yields along with minor amounts of their regioisomers (Scheme 13).



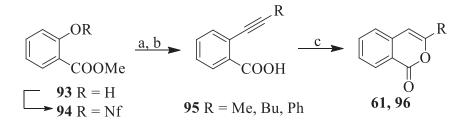


The total synthesis of naturally occurring dihydroisocoumarins such as hydrangenol, phyllodulcin, macrophyllol, and thunberginol G has been accomplished using titanocene(III) chloride (Cp_2TiCl) as a radical initiator.^{95,96} The Cp_2TiCl was prepared in situ from commercially available Cp_2TiCl_2 and Zn-dust. For example, ester **90** was brominated with NBS in the presence of the radical initiator AIBN yielding **91** in 92% yield. The bromo ester **91** afforded lactone **92** in 53% yield as a crystalline solid upon treatment with Cp_2TiCl in the presence of 4-methoxybenzaldehyde (Scheme 14).



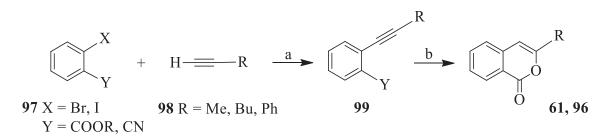
Reagent and conditions: a) NBS/AIBN, CCl₄; b) Cp₂TiCl₂/THF, 4-OMePhCHO.
Scheme 14. Synthesis of lactone 92.

Ogawa et al. reported a convenient method for the synthesis of isocoumarin derivatives via Ag-mediated intramolecular cyclization of 2-(1-alkynyl)benzoic acids **95**. The reaction first involves the formation of non-aflates **94** from **93**, followed by their Pd-catalyzed alkynylation, conversion of esters into the corresponding acid **95**, and Ag-salt-catalyzed 6-endo-dig cyclization of these acids that afforded isocoumarin **96**. The formation of side-product **68** is quite possible in this case due to 5-exo-dig ring closure. Some of the naturally occurring isocoumarin derivatives such as 3-propynylisocoumarin and attemidin were prepared using this strategy (Scheme 15).⁹⁷⁻⁹⁹



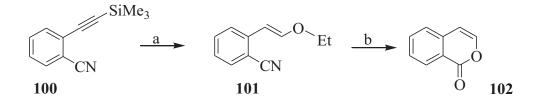
Reagent and conditions: a) terminal alkynes; b) NaOH; c) AgI or Ag, DMF.Scheme 15. Formation of isocoumarin 96 via Ag-mediated intramolecular cyclization.

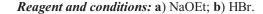
The 3-substituted isocoumarin derivatives 96 were prepared by coupling reaction of 97 and 98, followed by the hydration of 99 in the presence of HgSO₄ in H₂SO₄. The alkyne 99 was heated with HgSO₄ and dilute H₂SO₄ to afford isocoumarin derivative 96 with variable yields (Scheme 16). It was observed that alkynes from o-halobenzonitrile derivatives provide isocoumarin in poor yields; however, alkynes **100** are preferably used for the synthesis of unsubstituted isocoumarins **102** via alkene **101** (Scheme 17).^{100,101}



Reagent and conditions: a) PdCl₂(PPh₃)₃; b) HgSO₄, H₂SO₄.

Scheme 16. A preparation method for 3-substituted isocoumarin derivatives 96.



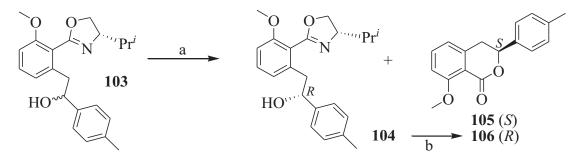


Scheme 17. Synthesis of unsubstituted isocoumarins 102 via alkene 101.

3.5. Asymmetric synthesis of isocoumarins and 3,4-dihydroisocoumarins

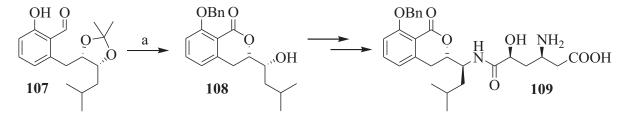
Iwao and coworkers¹⁰² devised a direct method for the synthesis of dihydroisocoumarin **105–106** by the reaction of oxazoline **103** and silica gel in CH_2Cl_2 at 0 °C via intermediate **104** (Scheme 18).

The AI-77s such as **109** are a group of 3,4-dihydroisocoumarin antibiotics that have been isolated from a culture broth of *Bacillus pumilus* AI-77.^{103–109} The AI-77-B **109** has been found to exhibit potent gastroprotective activity without anticholinergic, antihistaminergic, or central suppressive effects.^{110,111} The protection of **107** as its benzyl ether followed by deprotection of acetonide functionality yielded a diol, which was further oxidized by $NaClO_2/NaHSO_3$ and $30\% H_2O_2$ under carefully controlled conditions to afford lactone **108**. The dihydroisocoumarin **108** was then transformed to AI-77-B **109** (Scheme 19).¹¹²



Reagent and conditions: a & b) silica gel, CH₂Cl₂, 0 °C, 30 h.

Scheme 18. A direct method for the synthesis of dihydroisocoumarins 105-106 from oxazoline 103.

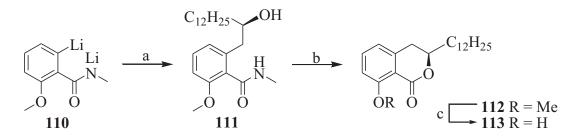


Reagent and conditions: a) i) BnBr, K₂CO₃; ii) HClO₄(cat.), CH₃CN; iii) NaClO₂, NaHSO₃, H₂O₂,

KHPO₄, aq. MeCN.

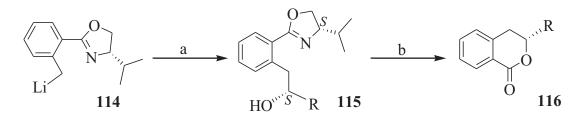
Scheme 19. A synthetic strategy for AI-77-B 109.

The enantiomerically pure 3,4-dihydroisocoumarins 112-113 have been obtained from lithiated secondary benzamides 110 and homochiral epoxides. The reaction proceeds through uncyclized intermediate 111. However, unfortunately, the yields are generally modest and N-alkylation can complicate the reaction.¹¹³ Good yields have occasionally been reported in a few cases such as in the syntheses of the antiallergic agent 113isolated from *Gingko biloba* (Scheme 20)¹¹⁴ and in the synthesis of a variety of mellein derivatives.¹¹⁵



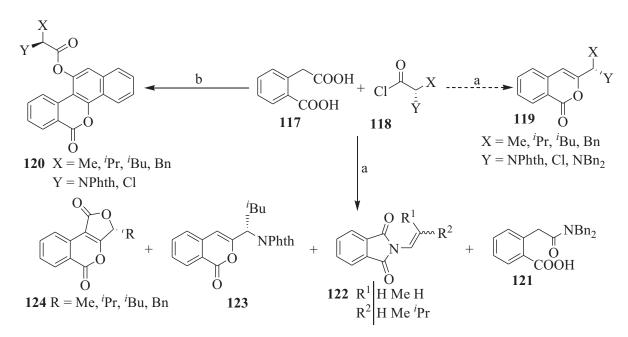
Reagent and conditions: a) (*R*)-1,2-epoxytetradecane; b) ⁻OH, neutralization CuCN(LiCl₂); c) BBr₃. Scheme 20. A scheme for the synthesis of enantiomerically pure 3,4-dihydroisocoumarins 112–113.

Lateral lithiation of (S)-4-isopropyl-2-(o-tolyl)oxazoline 114 in Et₂O followed by reaction with aldehydes in the presence of tetramethylethylenediamine (TMEDA) produced the major (S,S)-products 115 with high stereoselectivity (84% de).¹¹⁶ The adduct was then lactonized to the corresponding (3S)-3,4-dihydroisocoumarins **116** under acidic conditions in good optical purity (97% ee) (Scheme 21).



Reagent and conditions: a) RCHO; b) H₃O⁺ Scheme 21. Synthesis of dihydroisocoumarins 116 having chiral centers.

Saddiqa et al. reported the asymmetric synthesis of isocoumarins by the condensation of homophthalic acid **117** with different chiral carboxylic acids chlorides **118** at high (200 °C) and low (-5 °C) temperatures. The coupling at high temperature does not furnish **119**; instead, 3H-furo[3,4c] isochromene-1,11-diones **124** along with other side-products (**121**, **122**) are produced. Only the coupling reaction of phthaloyl *N*-protected leucine with homophthalic acid afforded **123** with poor yield (30%). The coupling at low temperature, in basic conditions, afforded chrysene-based (S)-isocoumarins **120** as a single product in high yields (Scheme 22).^{117,118}



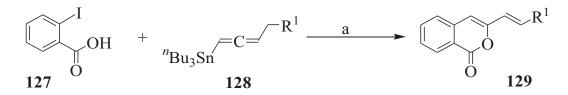
Reagent and conditions: a) 200 °C, neat b) i. SOCl₂, ii. Et₃N, -5 °C.

Scheme 22. The asymmetric synthesis of isocoumarins by the condensation of homophthalic acid 117 with different (S)-carboxylic acids chlorides 118.

3.6. Lewis acids-mediated cyclization

Bihel and coworkers¹¹⁹ synthesized 5-aza-3,4-dihydroisocoumarin **126** in excellent yields (up to 98%) via regiocontrolled 6-endo-dig cyclization of 2-(2-arylethynyl)heteroaryl ester **125**. The reaction was carried out

under microwave environment at 100 °C by employing a Bronsted acid in the presence of a catalytic amount of Lewis acids such as $Cu(OTf)_2$, $AuCl_3$, or $(CF_3CO_2)Ag$ (Scheme 23).

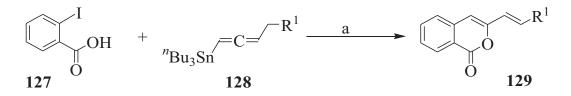


Reagent and conditions: a) Pd(OAc)₂ (5 mol%), PPh₃ (10 mol%), "Bu₄NBr (1 eq), DMF, 80 °C.

Scheme 23. Microwave-assisted synthesis of 5-aza-3,4-dihydroisocoumarin 126.

3.7. Synthesis of isocoumarins via tandem Stille coupling

A general route to 3-substituted isocoumarins **129** from 2-iodobenzoic acids **127** was described by Cherry et al.¹²⁰ The treatment of 2-iodobenzoic acids **127** with various allenyl-tri-n-butyltin reagents **128** in the presence of $Pd(OAc)_2$ [source of Pd(II)], PPh_3 (ligand), and Bu_4NBr (phase transfer reagent) in DMF provided good yields of the corresponding 3-substituted isocoumarins **129** via a tandem Stille reaction and 6-endo-dig oxacyclization (Scheme 24).



Reagent and conditions: a) Pd(OAc)₂ (5 mol%), PPh₃ (10 mol%), "Bu₄NBr (1 eq), DMF, 80 °C.

Scheme 24. A general route to 3-substituted isocoumarins 129 from 2-iodobenzoic acids 127 described by Cherry et al.¹²⁰

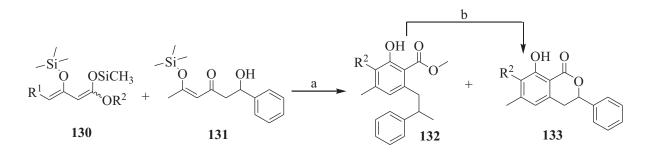
3.8. Regioselective cyclization of 1,3-bis(silyloxy)-1,3-butadienes

The [3+3] cyclization of 1,3-bis(silyloxy)-1,3-butadienes **130** with 1-hydroxy-5-silyloxy-hex-4-en-3-ones **131** resulted in the one-pot formation of 3-aryl-3,4-dihydroisocoumarins **133** (Scheme 25).¹²¹ The reactions proceeded by regioselective cyclization to give 6-(2-aryl-2-chloroethyl)salicylates **132**, which underwent a silical gel-mediated lactonization to afford lactones **133**.

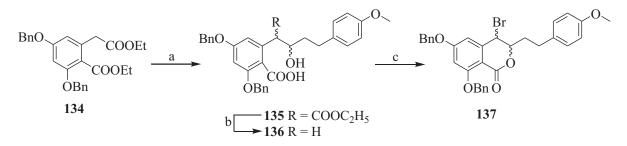
3.9. Aldol condensation

3.9.1. Stobbe's condensation

This type of condensation is mostly used in the synthesis of isocoumarins and 3,4-dihydroisocoumarins. Stobbe's condensation is used for the synthesis of a number of 3,4-dihydroisocoumarins.^{122–126} Synthesis of (dl)agrimonolide¹²⁷ provides a good example of application of Stobbe's condensation. Thus, homophthalate **134** upon condensation with 4-OMePhCHO in the presence of NaH afforded 2,4-dibenzyloxy-6-[1-ethoxycarbonyl-4-(4'-methoxyphenyl)buten-1-yl]benzoic acid **135** (R = COOEt). The hydrolysis and decarboxylation yielded 2,4-dibenzyloxy-6-[4-(4'-methoxyphenyl) buten-1-yl]benzoic acid **136** (R = H), which upon cyclization with Br_2 afforded 4-bromo-3,4-dihydroisocoumarin **137** (Scheme 26).

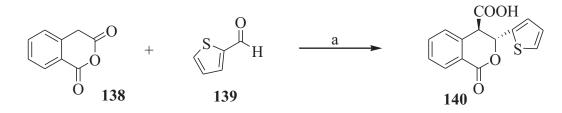


Reagent and conditions: a) i) TiCl₄, CH₂Cl₂ -78 °C, ii) NaHCO₃, H₂O; b) SiO₂ (wet), THF, 14 h. Scheme 25. A synthetic diagram for the lactone 133.



Reagent and conditions: **a**) 3-(4'-Methoxyphenyl)propanal; **b**) NaOH; **c**) Br₂, CHCl₃ Scheme 26. Synthesis of 4-bromo-3,4-dihydroisocoumarin 137 via Stobbe's condensation.

Bogdanov et al. carried out dimethylaminopyridine (DMAP)-assisted Stobbe's condensation of homophthalic anhydride **138** and thiophene-2-carbaldehyde **139** to afford 3-substituted trans-3,4-dihydroisocoumarin-4-carboxylic acids **140** (Scheme 27).¹²⁸

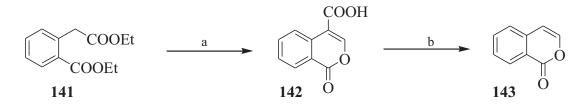


Reagent and conditions: a) DMAP, CHCl₃, rt.

Scheme 27. Stobbe's condensation reaction of homophthalic anhydride 138 and thiophene-2-carbaldehyde 139.

3.9.2. Claisen condensation of homophthalates with formates

The condensation of diethyl homophthalate **141** with methyl formate in the presence of NaOEt affords isocoumarin-4-carboxylic acid **142** at up to 66% yield. The decarboxylation of **142** with phosphoric acid furnishes isocoumarin **143** (Scheme 28).¹²⁹



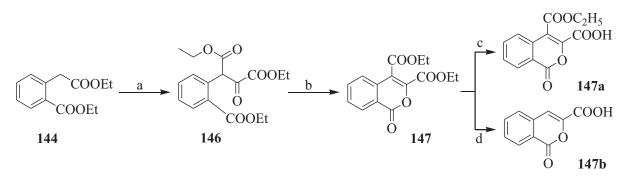
Reagent and coditions: a) HCOOCH₃, C₂H₅ONa; b) H₃PO₄, -CO₂

Scheme 28. A simple preparation method for the synthesis of isocoumarin 143.

The 6,7-dimethoxy isocoumarin and 5,7-dimethoxy isocoumarin were also prepared by the above procedure. The ethyl 5,6,7-trimethoxy isocoumarin-4-carboxy late was prepared from the corresponding homophthalate and ethyl formate in the presence of KOEt in good yield.¹³⁰

3.9.3. Claisen condensations of homophthalates with oxalates

The condensation between diethyl homophthalate 144 and diethyl oxalate 145 in the presence of Na in Et₂O, or better without a solvent, affords triester 146 in good yield (67%). This triester was heated, which yielded diethyl isocoumarin-3,4-dicarboxylate 147. Under different hydrolysis conditions, different products are formed. For example, heating 147 at 68–72 °C for 3 h furnishes ethyl isocoumarin-3-(carboxylic acid)-4-carboxylate 147a, and prolonged heating yields isocoumarin-3-carboxylic acid 147b. Boiling HCl or heating in a sealed tube at 180–190 °C converts 147 to isocoumarin-3-carboxylic acid in 84% yield.¹³¹ These results indicate that the ester at C³ in 147 is hydrolyzed first but the acid at position 4 is more easily decarboxylated (Scheme 29).

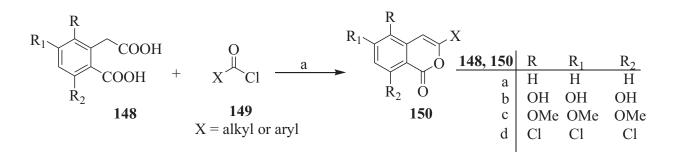


Reagent and conditions: a) diethyl oxalate 145, Na; b) Δ ; c) 68–72 °C; d) prolong heating. Scheme 29. Synthesis of isocoumarins by Claisen condensations of homophthalates and oxalates.

3.9.4. Condensation of acid chlorides with homophthalic acids and anhydrides

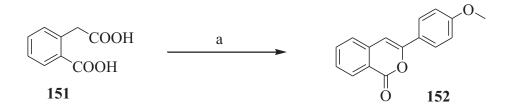
Nakajima et. al. synthesized various 3-arylisocoumarins **150** and later on 3-alkyl isocoumarins in high yields (80%) by directly heating the homophthalic acids **148** with aryl or acyl chlorides **149**. These isocoumarins were converted into corresponding 3,4-dihydroisocoumarins by reduction with NaBH₄ (Scheme 30).¹³¹

The 3-(4'-methoxy phenyl)isocoumarin ${\bf 152}$ was prepared by condensation of homophthalic acid ${\bf 151}$ with anisole (Scheme 31). 132



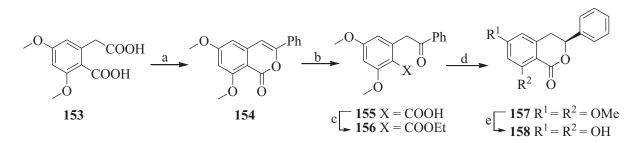
Reagent and conditions: a) 200 °C, 6 h.

Scheme 30. Condensation of acid chlorides 149 with homophthalic acids.



Reagent and conditions: a) anisole, PPA (polyphthalamide), rt. Scheme 31. The synthetic route for 3-(4'-methoxyphenyl)isocoumarin 152.

The 6,8-dimethoxy-3-phenylisocoumarin **154** was prepared by condensation of 3,5-dimethoxyhomophthalic acid **153** with benzoyl chloride at 200 °C. The isocoumarin was hydrolyzed to **155** and esterified to furnish ketoester **156** that was further enantioselectively reduced to afford (3S)-6,8-dimethoxy-3-phenyl-3,4-dihydroisocoumarin **157**. The demethylation of **157** afforded (3S)-6,8-dihydroxy-3-phenyl-3,4-dihydroisocoumarin **158** (Scheme 32).¹³³



Reagent and conditions: a) PhCOCl, 200 °C, 4 h; b) 5% KOH, EtOH, 4 h, reflux; c) CH₃I, K₂CO₃, dry

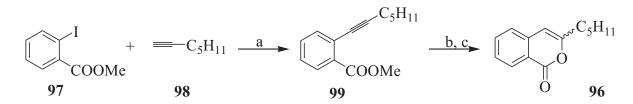
acetone, 5 h; d) Baker's yeast; e) BBr₃, CH₂Cl₂, -78 °C, overnight.

Scheme 32. Preparation of 3-phenyl-3,4-dihydroisocoumarins 157 and 158.

3.10. Cyclization of methyl 2-heptynylbenzoate

Villemin et al. reported the synthesis of isocoumarin **96** by the coupling of o-iodobenzoic ester **97** and a terminal alkyne **98**, catalyzed by Pd salt and Cu-catalyst. The reaction proceeds under Sonogashira conditions

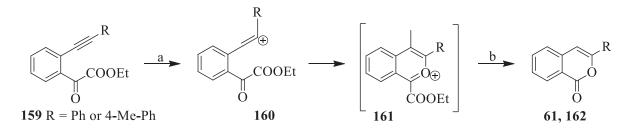
and yields 2-alkynyl benzoic ester 99, which upon successive saponification and acidification gave isocoumarin 96 as a major product (Scheme 33).¹³⁴



Reagent and conditions: a) L₂PdCl₂.Cu₂I₂, Et₃N; b) KOH; c) H₂SO₄.
Scheme 33. Cyclization reaction of 99 and a terminal alkyne 98.

3.11. Synthesis via isobenzopyrylium salts

Pyrylium salts play very important roles in organic synthesis as they are useful intermediates for the synthesis of many heterocyclic nuclei and have also been used for the synthesis of different derivatives of isocoumarins. The alkyne **159** was prepared by modified Sonogashira procedure from o-iodobenzoic ester. These esters undergo quantitative cyclization in the presence of strong acids such as HBF₄ and TfOH to give salt **161** via **160**, which are unstable. The slow hydrolysis of the tetrafluoroborate salts **161** at room temperature yielded the isocoumarin **162** (Scheme 34).¹³⁵



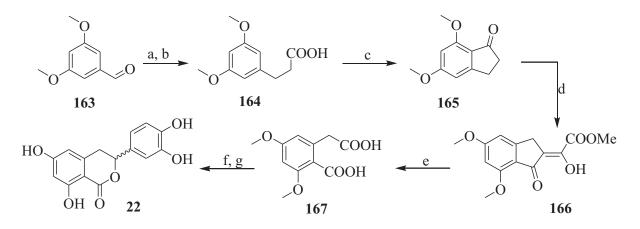
Reagent and conditions: a) HX, [X=BF₄ or TfO], CH₂Cl₂; b) hydrolysis. Scheme 34. Synthesis of isocoumarin 61, 162 via isobenzopyrylium salt 161.

3.12. Synthesis of naturally occurring isocoumarin derivatives

The synthesis of a number of naturally occurring isocoumarins is available in literature; for example, Qadeer et al. reported the synthesis of thunberginol B **22** by the coupling of 3,5-dimethoxy homophthalic acid **167** with 3,4-dimethoxybenzoic acid followed by the demethylation of the intermediate. The 3,5-dimethoxyhomophthalic acid **167** was synthesized in five steps, starting from 3,5-dimethoxybenzaldehyde **163**, and the reaction proceeded through **164–166** (Scheme 35).¹³⁶

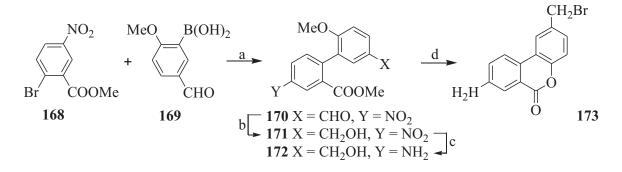
3.13. Synthesis of pharmacologically active isocoumarin derivatives

A number of compounds having an isocoumarin nucleus are found as inhibitors of various enzymes such as serine proteases, HIV aspartyl protease, and a panel of protein kinases, e.g., 2,8-disubstituted-benzo[c]chromen-6-ones. The Suzuki coupling of bromoarene **168** with boronic acid derivative **169** afforded the biaryl compound **170**, which upon successive reduction of the CHO and NO₂ group yielded the ester **172** via **171**. The cyclization of **172** provided the required isocoumarin **173** (Scheme 36).¹³⁷



Reagent and conditions: a) CH₂(COOH)₂; b) Na/Hg; c) PPA; d) (CO₂Et)₂, NaOMe; e) H₂O₂, KOH; f) 3,4-dimethoxybenzoyl chloride, 200 °C, reflux; g) HBr.

Scheme 35. Total synthesis of thunberginol B 22.



Reagent and conditions: a) i. PdCl₂ dppf; ii. dppf, KOAc, dioxane, reflux; b) B₂H₆/DMS, THF, rt, 1 h;

c) H₂/Pd (C), THF, rt, 1 h; d) BBr₃, CH₂Cl₂, -78 °C, 2 h, CH₃OH.

Scheme 36. Synthesis of pharmacologically active isocoumarin 173.

4. Conclusion

The isocoumarin and 3,4-dihydroisocoumarins ring system is found in nature with a wide spectrum of biological activities, ranging from antibacterial to anticancer. Based on this review, it can be concluded that due to distinctive pharmacological significance of these motifs, much research has been done and still going on towards the development and synthesis of their derivatives.

References

- 1. Filho, R. B.; De Moraes, M. P. L.; Gottieb, O. R. Phytochemistry 1980, 19, 2003-2006.
- 2. Hill, R. A. Prog. Chem. Org. Nat. Prod. 1986, 49, 1-78.
- 3. Zepnik, H.; Pahler, A.; Schauer, U.; Dekant, W. Toxicol. Sci. 2001, 59, 59-67.
- 4. Barry, R. D. Chem. Rev. 1964, 64, 229-260.
- 5. Turner, W. B.; Aldridge, D. C. Fungal Metabolites II; Academic Press: London, UK, 1983.

- 6. Napolitano, E. Org. Prep. Proced. Int. 1997, 29, 631-664.
- 7. Bin, Y.; Song, L.; Xiaohui, G. Tianran Chanwu Yanjiu Yu Kaifa 2000, 12, 95-98.
- 8. Saeed, A. Eur. J. Med. Chem. 2016, 116, 290-317.
- 9. Vogel, A. Ann. Phys. 1820, 64, 161-166.
- 10. Pochet, L.; Frederick, R.; Masereel, B. Curr. Pharm. Des. 2004, 10, 3781-3796.
- 11. Chen, H. W.; Walsh, C. T. Novel Chem. Biol. 2001, 8, 301-312.
- 12. Ito, J.; Omoto, S.; Shomura, T.; Nishizawa, N.; Miyado, S.; Yuda, Y.; Shibata, U.; Inoue, S. J. Antibiot. 1981, 34, 611-613.
- 13. Patel, S. K.; Murat, K.; Py, S.; Vallee, Y. Org. Lett. 2003, 5, 4081-4084.
- 14. Okazaki, H.; Kishi, T.; Beppu, T.; Arima, K. J. Antibiot. 1975, 28, 717-719.
- 15. Weisenborn, F. L.; Brown, W. E.; Meyers, E. US Patent 4296101, 1981.
- 16. Canedo, L. M.; Fernandez-Puentes, J. L.; Perez-Baz, J.; Acebal, C.; De la-Calle, F.; Garcia, G. D.; Garcia, T. J. Antibiot. 1997, 50, 175-176.
- Sato, T.; Nagai, K.; Suzuki, K.; Morioka, M.; Saito, T.; Nohara, C.; Susaki, K.; Takebayashi, Y. J. Antibiot. 1992, 45, 1949-1952.
- 18. Hernandez, L.; Canedo, S.; Acebal, G. C.; Garcia, D. US Patent 5925671, 1999.
- 19. Huang, Y. F.; Li, L. H.; Tian, L.; Qiao, L.; Hua, H. M.; Pei, Y. H. J. Antibiot. 2006, 59, 355-357.
- 20. Stefano, S.; Filippo, M.; Dario, P.; Piero, S. J. Org. Chem. 1996, 61, 3183-3186.
- 21. Shimojima, Y.; Hayashi, H.; Ooka, T.; Shibukawa, M.; Iitaka, Y. Tetrahedron Lett. 1982, 23, 5435-5438.
- 22. Shimojima, Y.; Hayashi, H.; Ooka, T.; Shibukawa, M. Tetrahedron 1984, 40, 2519-2527.
- Ito, J.; Shomura, T.; Omoto, S.; Miyado, S.; Yuda, Y.; Shibata, U.; Inoue, S. Agric. Biol. Chem. 1982, 46, 1255-1259.
- 24. McInerney, B. V.; Taylor, W. C.; Lacey, M. J.; Akhurst, R. J.; Gergson, R. P. J. Nat. Prod. 1991, 54, 785-795.
- 25. Hiyoshizo, K.; Tomohiro, A.; Aya, A.; Mitsuhiro, I.; Probal, K. D. Org. Lett. 1999, 1, 499-502.
- 26. Simon, D.; Broady, J.; Rexhausen, E.; Thomas, E. J. J. Chem. Soc. Perk. T. 1, 1999, 8, 1083-1094.
- 27. Vilcinskas, A. Insect Biotechnology; Springer: New York, NY, USA, 2010.
- 28. Shimojima, Y.; Shirai, T.; Baba, T.; Hayashi, H. J. Med. Chem. 1985, 28, 3-9.
- 29. Shimojima, Y.; Hayashi, H. J. Med. Chem. 1983, 26, 1370-1374.
- McInerney, B. V.; Taylor, W. C. In *Studies in Natural Products Chemistry*; Atta-ur-Rahman, Ed.; Elsevier Science: Amsterdam, the Netherlands, 1995, pp. 381-422.
- 31. Hill, R. A. Prog. Chem. Org. Nat. Prod. 1986, 49, 1-78.
- 32. Piacente, S.; Pizza, C.; De Tommasi, N.; Mahmood, N. J. Nat. Prod. 1996, 59, 565-569.
- 33. Singh, I. P.; Bharate, S. B.; Bhutani, K. K. Curr. Sci. 2005, 89, 269-290.
- 34. Goel, R. K.; Maiti, R. N.; Manickam, M.; Ray, A. B. Ind. J. Exp. Biol. 1997, 35, 1080-1083.
- 35. Kim. S. H. J. Ethnopharmacol. 2000, 69, 79-83.
- 36. Pu, H. L.; Huang, X.; Zhao, J. H.; Hong, A. Planta Med. 2002, 68, 372-374.
- 37. Prithiviraj, B.; Singh, U. P.; Manickam, M.; Srivastava, J. S.; Ray, A. B. Plant Pathol. 1997, 46, 224-228.
- 38. Devienne, K. F.; Raddi, M. S. G.; Varanda, E. A.; Vilegas, W. Z. Naturforsch. 2002, 57c, 85-88.
- 39. Rama, N. H.; Hussain, S. J. Arkivoc 2007, 15, 12-19.
- 40. Lee, J. J.; Kim, H. S.; Lee, J. H.; Hong, Y. S.; Park, Y. J. US Patent 6,451,846 B1, 2002.
- 41. Buckle, D. R.; Cantello, B. C. C.; Smith, H. US Patent 3,975,535, 1976.

- 42. Power, J. C.; Kam, C. M.; Oleksyszyn, J.; Glinski, J. A.; Hernandez, M. A. US Patent 5,324,648, 1994.
- 43. Heynekamp, J. J.; Hunsaker, L. A.; Jagt, T. A. V.; Deck, L. M. BMC Chem. Biol. 2006, 6, 1-11.
- 44. Kam, C. M.; Fujikawa, K.; Powers, J. C. Biochemistry 1988, 27, 2547-2557.
- 45. Rama, N. H.; Iqbal, R.; Rauf. A.; Zamani, K.; Raza, A. R. Ind. J. Chem. 1998, 37B, 338-341.
- 46. Hussain, M.; Hussain, M. T.; Rama, N. H.; Hameed, S.; Malik, A.; Khan, K. M. Nat. Prod. Res. 2002, 17, 207-214.
- 47. Saeed, A. J. Chin. Chem. Soc. 2003, 50, 313-317.
- Arunpanichlert, J.; Rukachaisirikul, V.; Phongpaichit, S.; Sukpondma, Y.; Sakayaroj, J. Tetrahedron 2009, 65, 10590-10595.
- 49. Saeed, A. Eur. J. Chem. 2011, 2, 117-119.
- Kihampa, C.; Nkunya, M. H. H.; Joseph, C. C.; Magesa, S. M.; Hassanali, A.; Heydenreich, M.; Kleinpeter, E. Phytochemistry 2009, 70, 1233-1238.
- Chinworrungsee, M.; Kittakoop, P.; Isaka, M.; Chanphen, R.; Tanticharoen, M.; Thebtaranonth, Y. J. Chem. Soc. Perk. T. 1 2002, 1, 2473-2476.
- 52. Tabacchi, R. Pure Appl. Chem. 1994, 66, 2299-2302.
- 53. Claydon, N.; Grove, J. F.; Hosken, M. Phytochemistry 1974, 13, 2567-2571.
- 54. Varanda, E. A.; Devienne, K. F.; Raddi, M. S. G.; Furuya, E. M.; Vilegas, W. Toxicol. In Vitro 2004, 18, 109-114.
- 55. Okuno, T; Oikawa, S; Sawai, K; Shirahama, H.; Matsumoto, T. Agric. Biol. Chem. 1986, 50, 997-1001.
- 56. Rama, N. H.; Hussain, M.; Hussain, M. T.; Hameed, S.; Malik, A. Nat. Prod. Res. 2002, 17, 207-214.
- 57. Zhang, W.; Krohn, K.; Draeger, S.; Schulz, B. J. Nat. Prod. 2008, 71, 1078-1081.
- 58. Ozcan, S.; Balci. M. Tetrahedron 2008, 64, 5531-5540.
- 59. Munakata, T.; Okumoto, T. Chem. Pharm. Bull. 1981, 29, 891-894.
- 60. Hirano, S. I.; Mase, T.; Agata, N.; Iguchi, H.; Kumagai, H. US Patent 6,020,363A, 2000.
- 61. Bestman, H. J.; Kern, F.; Schafe, D.; Witschel, M. C. Angew. Chem. Int. Edit. 1992, 31, 795-796.
- 62. Watanabe, W.; Sahara, M.; Furukawa, S.; Billedeau, R.; Snieckus, V. Tetrahedron Lett. 1982, 23, 1647-1650.
- 63. Barry, R. P.; Chem. Rev. 1964, 64, 229-260.
- 64. Mali, R. S.; Babu, K. N. J.Org. Chem. 1998, 63, 2488-2492.
- Zidorn, C.; Lohwasser, U.; Pschorr, S.; Salvenmoser, D.; Ongania, K. H.; Ellmerer, E. P.; Börner, A.; Stuppner, H. Phytochemistry 2005, 66, 1691-1697.
- Umehara, K.; Matsumoto, M.; Nakamura, M.; Miyase, T.; Kuroyanagi, M.; Noguchi, H. Chem. Pharm. Bull. 2000, 48, 566-567.
- 67. Qin, D.; Ren, R. X.; Siu, T.; Zheng, C.; Danishefsky, S. J. Angew. Chem. Int. Edit. 2001, 40, 4709-4713.
- 68. Siu, T.; Qin, D.; Danishefsky, S. J. Angew. Chem. Int. Edit. 2001, 40, 4713-4716.
- 69. Narasimhan, N. S.; Mali, R. S. Top. Curr. Chem. 1987, 138, 63-147.
- 70. Woon, E. C. Y.; Dhami, A.; Mahon, M. F.; Threadgill, M. D. Tetrahedron 2006, 62, 4829-4837.
- 71. Subramanian, V.; Batchu, V. R.; Barange, D.; Pal, M. J. Org. Chem. 2005, 70, 4778-4783.
- 72. Roy, H.; Sarkar, M. Synth. Commun. 2005, 35, 2177-2181.
- 73. Cherry, K.; Parrain, J. L.; Thibonnet, J.; Duchene, A.; Abarbri, M. J. Org. Chem. 2005, 70, 6669-6675.
- 74. Suzuki, T.; Yamada, T.; Watanabe, K.; Katoh, T. Bioorg. Med. Chem. Lett. 2005, 15, 2583-2585.
- 75. Opatz, T.; Ferenc, D. Eur. J. Org. Chem. 2005, 5, 817-821.
- 76. Martinez, A.; Fernandez, M.; Estevez, J. C.; Estevez, R. J.; Castedo, L. Tetrahedron 2005, 61, 485-492.
- 77. Yao, T.; Larock, R. C. J. Org. Chem. 2003, 68, 5936-5942.

- 78. Liao, H. Y.; Cheng, C. H. J. Org. Chem. 1995, 60, 3711-3716.
- 79. Hussain, M.; Rama, N. H.; Hameed, S.; Malik, A.; Khan, K. M. Nat. Prod. Res. 2005, 19, 41-51.
- 80. Zamani, K.; Faghihi, K.; Ebrahimi, S. Turk. J. Chem. 2005, 29, 171-175.
- 81. Hauser, F. M.; Baghdanov, V. M. J. Org. Chem. 1988, 53, 4676-4681.
- Uchiyama, M.; Ozawa, H.; Takuma, K.; Matsumoto, Y.; Yonehara, M.; Hiroya, K.; Sakamoto, T. Org. Lett. 2006, 8, 5517-5520.
- Yoshikawa, M.; Matsuda, H.; Shimoda, H.; Shimada, H.; Harada, E.; Naitoh, Y.; Miki, A.; Yamahara, J.; Murakami, N. Chem. Pharm. Bull. 1996, 44, 1440-1447.
- 84. Yoshikawa, M.; Harada, E.; Naitoh, Y.; Inoue, K.; Matsuda, H.; Shimoda, H.; Yamahara, J.; Murakami, N. Chem. Pharm. Bull. 1994, 42, 2225-2230.
- 85. Yoshikawa, M.; Uchida, E.; Chatani, N.; Kobayashi, H.; Naitoh, Y. Chem. Pharm. Bull. 1992, 40, 3352-3354.
- 86. Yoshikawa, M.; Uchida, E.; Chatani, N.; Murakami, N.; Yamahara, J. Chem. Pharm. Bull. 1992, 40, 3121-3123.
- 87. Menashe, N.; Shvo, Y. Heterocycles 1993, 35, 611-613.
- 88. Castro, C. E.; Stephens, R. D. J. Org. Chem. 1963, 28, 3313-3315.
- 89. Sharma, A. K.; Maheshwary, Y.; Singh, P.; Singh, K. N. Arkivoc 2010, 2010, 54-62.
- 90. Deshpande, V. H.; Rai, B.; Khan, R. A. Tetrahedron 1996, 52, 7159-7162.
- 91. Conners, R.; Tran, E.; Durst, T. Can. J. Chem. 1996, 74, 221-226.
- 92. Kawasaki, T.; Saito, S.; Yamamoto, Y. J. Org. Chem. 2002, 67, 2653-2658.
- 93. Suzuki, T.; Yamada, T.; Watanabe, K.; Katoh, T. Biorg. Med. Chem Lett. 2005, 15, 2583-2585.
- 94. Marchal, E.; Uriac, P.; Legouin, B.; Toupet, L.; Van de Weghe, P. Tetrahedron 2007, 63, 9979-9990.
- 95. Ueura, K.; Satoh, T.; Miura, M. J. Org. Chem. 2007, 72, 5362-5367.
- 96. Mandal, S. K.; Roy, S. C. Tetrahedron Lett. 2007, 48, 4131-4234.
- 97. Mandal, S. K.; Roy, S. C. Tetrahedron 2008, 64, 11050-11057.
- 98. Bellina, F.; Ciucci, D.; Vergamini, P.; Rossi, R. Tetrahedron 2000, 56, 2533-2545.
- 99. Ogawa, Y.; Maruno, M.; Wakamatsu, T. Synlett 1995, 1995, 871-873.
- 100. Ogawa, Y.; Maruno, M.; Wakamatsu, T. Heterocycles 1995, 41, 2587-2599.
- 101. Sakamoto, T.; An-Naka, M.; Kondo, Y.; Yamanaka, H. Chem. Pharm. Bull. 1986, 34, 2754-2759.
- 102. Nagarajan, A.; Balasubramanian, T. R. Ind. J. Chem. Sect. B, 1987, 26, 917-919.
- 103. Uchida, K.; Fukuda, T.; Iwao, M. Tetrahedron 2007, 63, 7178-7186.
- 104. Shimojima, Y.; Hayashi, H.; Ooka, T.; Shibukawa, M. Agric. Biol. Chem. 1982, 46, 1823-1829.
- 105. Shimojima, Y.; Hayashi, H.; Ooka, T.; Shibukawa, M.; Iitaka, Y. Tetrahedron Lett. 1982, 23, 5435-5438.
- 106. Shimojima, Y.; Hayashi, H.; Ooka, T.; Shibukawa, M.; Iitaka, Y. Tetrahedron 1984, 40, 2519-2527.
- 107. Okazaki, H.; Kishi, T.; Beppu, T.; Arima, K. J. Antibiot. 1975, 28, 717-719.
- 108. Itoh, J.; Omoto, S.; Shomura, T.; Nishizawa, N.; Miyado, S.; Yuda, Y.; Shibata, U.; Inoue, S. J. Antibiot. 1981, 34, 611-613.
- 109. Itoh, J.; Shomura, T.; Omoto, S.; Miyado, S.; Yuda, Y.; Shibata, U.; Inouye, S. Agric. Biol. Chem. 1982, 46, 1255-1259.
- 110. Itoh, J.; Omoto, S.; Nishizawa, N.; Kodama, Y.; Inouye, S. Agric. Biol. Chem. 1982, 46, 2659-2665.
- 111. Shimojima, Y.; Hayashi, H. J. Med. Chem. 1983, 26, 1370-1374.
- 112. Shimojima, Y.; Shirai, T.; Baba, T.; Hayashi, H. J. Med. Chem. 1985, 28, 3-9.
- 113. Kotsuki, H.; Araki, T.; Miyazaki, A.; Iwasaki, M.; Datta, P. K. Org. Lett. 1999, 1, 499-502.

- 114. Choukchou-Braham, N.; Asakawa, Y.; Lepoittevin, J. P. Tetrahedron Lett. 1994, 35, 3949-3952.
- 115. Bhide, B. H.; Akolkar, V. D.; Brahmbhat, D. I. Ind. J. Chem. 1992, 31B, 116-117.
- 116. Kurosaki, Y.; Fukuda, T.; Iwao, M. Tetrahedron 2005, 61, 3289-3303.
- 117. Mills, R. J.; Taylor, N. J.; Sieckus, V. J. Org. Chem. 1989, 54, 4372-4385.
- 118. Saddiqa, A.; Raza, A. R.; Black, D. S. C; Kumar, N. Tetrahedron Asymmetry 2014, 25, 736-743.
- 119. Raza, A. R.; Saddiqa, A.; Çakmak, O. Chirality 2015, 27, 951-957.
- 120. Hellal, M.; Bourguignon, J. J.; Bihel, F. J. J. Tetrahedron Lett. 2008, 49, 62-65.
- 121. Cherry, K.; Parrain, J. L.; Thibonnet, J.; Duchêne, A.; Abarbri, M. J. Org. Chem. 2005, 70, 6669-6675.
- 122. Ullah, I.; Sher, M.; Khera, R. A.; Ali, A.; Ibad, M. F.; Villinger, A.; Fischer, C.; Langer, P. Tetrahedron 2010, 66, 1874-1884.
- 123. Loewenthal, H. J. E.; Pappo, R. J. Chem. Soc. 1952, 4799-4804.
- 124. Chatterjee, J. N.; Mukherjee, H. Experietia 1960, 16, 439-440.
- 125. Chatterjee, J. N.; Mukherjee, H. J. Ind. Chem. Soc. 1960, 37, 379-391.
- 126. Chatterjee, J. N.; Mukherjee, H. J. Ind Chem. Soc. 1960, 37, 443-450.
- 127. Yamato, M.; Hashigaki, K. Chem. Pharm. Bull. 1976, 24, 200-203.
- 128. Kabayashi, T. Sci. Rept. 1942, 31, 73-85.
- 129. Bogdanov, M.; Kandinska, M.; Yliev, B.; Palamareva, M. Pharmacia 2005, 2, 7-11.
- 130. Vorozhtsov, N. N.; Petushova, A. T. J. Gen. Chem. USSR 1957, 27, 2282-2284.
- 131. Chatterjee, J. N. J. Ind. Chem. Soc. 1953, 30, 103-112.
- 132. Kaji, H.; Yamada, M.; Nozawa, K.; Kawai, K. I.; Nakajima, S. Org. Prep. Proceed. Int. 1986, 1, 253-262.
- 133. Tuanli, Yao.; Richard, C. L. Tetrahedron Lett. 2002, 43, 7401-7404.
- 134. Saeed, A. Z. Naturforsch. 2003, 58c, 691-686.
- 135. Villemin, D.; Goussu, D. Heterocycle 1989, 29, 1255-1261.
- 136. Tovar, J. D.; Swager, T. M. J. Org. Chem. 1999, 64, 6499-6504.
- 137. Qadeer, G.; Rama, N. H.; Shah, J. H. Arkivoc 2007, 14, 12-19.
- 138. Garino, C.; Bihel, F.; Pietrancosta, N.; Laras, Y.; Quelever, G.; Woo, I.; Klein, P.; Bain, J.; Boucherd, J. L.; Kraus, J. L. *Bioorg. Med. Chem. Lett.* 2005, 15, 135-138.